Cat No: PPS-M-10X-500 Size: 500ml Stored at 18-25°C

**MQ Ponceau S Protein Staining Solution (10X)** 

#### Description

MQ Ponceaus S Staining Solution,10X concentrate is a membrane stain for evaluating the transfer efficiency of a western blot. This stain is recommended for rapid and reversible protein staining on nitrocellulose or PVDF membranes. Ponceau S staining is reversible and can be removed with a short incubation in 0.1% NaOH.



1 2 3 4 5 6 7 8 9 10 11 12 13

Ponceau S solution can be used to evaluate for total protein amount or transfer efficiency on nitrocellulose and PVDF membrane. The PVDF membrane stained Ponceau S for 5 mins and washed with ddH<sub>2</sub>O for 3 mins. It provides visible pink bands.

Lane 1, 13: Prestained MQ Protein Ladder Lane 2-7: 2X Dilutions of Unstained Protein Ladder Lane 8-12: 2000, 1000, 500, 100, 50 ng of BSA

## **Required materials but not provided**

- Container: box for gel staining
- Shaker: orbital or rocking shaker

## **Reagent Dilution**

- 1. Prepare a 5L amber bottle.
- 2. Unpack PPS-M-10X-500 and pour the liquid content into the bottle.
- 3. Rinse residual reagent in the original bottle with deionized water.
- 4. Add deionized water until the volume reaches 5L.

# **Reaction Setup**

- 1. Staining Solution. Make 10X concentrate MQ Ponceau S Protein Staining Solution into 1X concentrate MQ Ponceau S Protein.
- 2. After the SDS-PAGE Gel has been transferred to a nitrocellulose or PVDF membrane, place the transferred membrane in a staining container.
- 3. Submerge the membrane in a proper amount of 1X MQ Ponceau S Protein Staining Solution that is enough to cover the whole membrane. Lightly agitate the staining container for 5 to 15 minutes at room temperature. Protein bands will be stained and start becoming visible in 5 to 15 minutes.
- 4. Wash away the excessive stain on the membrane with deionized water until the protein bands are clear.
- 5. For the further membrane blocking step, remove the MQ Ponceau S Staining Solution from the membrane by placing the membrane in 20 ml 0.1% NaOH for 1 to 2 minutes then rinse in deionized water for 1 to 2 minutes.

## For Laboratory Use Only